

In vitro maintenance of bioactive Ca2+ and primary metabolite status in Cissus quadrangularis under salt stress

Renu Rani and Y Vimala*

Department of Botany, Ch. Charan Singh University, Meerut-250004, Uttar Pradesh, India

Received for publication: January 05, 2013; Accepted: February 28, 2013.

Abstract: Cissus quadrangularis is a plant, which has been customarily used in the Indian subcontinent to hasten the process of healing in bone fracture. The production of callus tissue was achieved from mature nodal segments by culturing on MS medium supplemented with auxin and cytokinin. Addition of 50mM salt to the medium, decreased potassium and phosphorus content of callus, but increased nitrogen, sodium and calcium content. Further increase in NaCl concentration decreased phosphorus, but increased nitrogen, potassium, sodium and calcium content as compared to control (without salt). Accumulation of calcium ions at higher salt concentration may possibly help in faster remodeling of bones. Calcium ion also mitigates the adverse effects of salinity up to a certain level.

Keywords: Bioactive Ca²⁺, Cissus quadrangularis, NaCl, Salt overly sensitive.

Introduction

Plants are not incompatible to salts, but most of the plants do not grow in saline soils. Globally, soil salinity is more common in arid and semi-arid regions than in humid regions. An understanding of responses of plants to salinity is of great practical significance. High concentrations of salts have detrimental effects on plant growth (1; 2). Many investigators have reported retardation of germination and growth of seedlings at high salinity (3; 1; 4). However, plant species differ in their sensitivity or tolerance to salts. There are many different types of salts and almost an equally diverse set of mechanisms of avoidance or tolerance. In addition, organs, tissues and cells at different developmental stages of plants exhibit varying degrees of tolerance to environmental conditions (5). Cissus quadrangularis L. (Vitaceae) commonly known as Hadjod is an important medicinal plant of great economic value (6). The species of Vitaceae found in India are edible, ornamental and have great medicinal properties including anthelmintic, alterative, dyspeptic, antiscorbic, stomachic, antioxidant and antimicrobial (7; 8; 9). It is a rambling shrub usually found in hotter parts of India, Sri Lanka, Malaysia, Java and West Asia. The stem is thick, fleshy, glabrous, and ridged and quadranglular at the nodes. It is referred as "Asthisamhari" in Sanskrit, "hadjod" in Hindi because of its ability to join bones and is commonly known as the "bone setter" in English. It is an edible plant; the stems are made into curries, chutneys and eaten in Southern parts of India.

The ash of the plant is being used as substitute for baking powder. The stem is useful in piles. The juice of the plant is used to control irregular menstruation, diseases of the ear and in nose- bleeding. A paste of the stem is given in asthma and may be useful for muscular pains, burns, and wounds. A decoction of the shoots with dry ginger and black pepper is given for body pain. The infusion of the plant is anthelmintic. The entire plant is being used in fractures, sprains, rheumatism and irregular growth of teeth, broken horn, anthrax, haematuria and dislocation of hip, various elephantiasis, The present wounds and cracked tail. investigation was carried out understand the adaptive features of Cissus quadrangularis which allow it to grow and survive in saline and arid regions, and (ii) to assess the pattern of macro- and micronutrient status within the tissues of this species in response to salt stress.

Material and Methods

Healthy plants of Cissus. collected from *quadrangularis* were botanical garden, Department of Botany, C.C.S.U. Campus Meerut. The nodal segments were used as explants for callus initiation. These explants were cut 8-10 mm and washed under running tap water for half an hour, subsequently followed by 1% labolene (Qualigens, India) for 5 min and then washed with sterile distilled water. They were then surface sterilised with 0.1% HgCl₂ solution for 3-5 min followed by several rinses in sterile

Prof. Y. Vimala Department of Botany C.C.S. University Meerut-250004, U.P., INDIA. distilled water. The sterile explants were then transferred on MS medium (10). The medium was supplemented with various combinations of auxins and cytokinins for callus induction. The media were congealed with agar (0.8%) and sucrose (3%) was used as a source of carbohydrate. pH of the medium was adjusted to 5.8 using 0.1 N NaOH/0.1N HCl solutions before being autoclaved at 15 psi for 20 min. All the cultures were incubated at 30±2°C under 16 h photoperiod, illuminated by fluorescent light of about 2500-3000 lux intensity. Biochemical parameters Nitrogen content (11), Phosphorus content (12), Mineral components (13) were analysed after six week of callus induction. All the experiments and measurements were made in the triplicate.

Results and Discussion

Callus growth was good to moderate in 2.5-3.0 mg/l NAA+0.5mg/l BA. Addition of 50mM NaCl led to increase in N (Fig. 1), Na⁺, and Ca²⁺ but decline in growth (Table 1), P (Fig. 2) and K⁺ content in the 6 week old calli. As the growth of callus is also reduced as compared to control, the primary metabolism may be considered to be under salt stress. The stress further enhanced by addition of 100mM NaCl to the basal MS medium. The energy metabolites (primary phosphorylating metabolite) reduced due to lower uptake of P in the presence of NaCl. Yet the interesting feature increased uptake was accumulation of Ca²⁺ (Fig. 3) which in turn induces the SOS (Salt Overly Sensitive) pathway (14) for exclusion of Na⁺. However, decline in K⁺ (Fig. 4) at lower NaCl concentration under 2.5mg/l NAA+0.5mg/l BA excludes the possibility of using the PGR combination supplementation for growing salt stressed calli vielding high amount of bioactive principle. Using the 3.0ma/l NAA+0.5mg/l BA resulted in improvement in accumulation of P, K⁺ and Ca²⁺ under 100mM NaCl along with moderately good growth of calli. This suggests that induction of SOS pathway by Ca²⁺ and also improvement of binding and uptake of NAA (15) under as low as 4.0mM Ca²⁺ which is provided in the standard nutrient composition of MS medium, helps in combating stress through increase in P uptake (though less than control) and K⁺ accumulation along with maintenance of balance of Na⁺ content equal to controls (Fig. 5). In vitro salt stress study in a plant like Cissus quadrangularis is suggestive of bioactive Ca²⁺ accumulation only at higher

salt concentration given to at least 3.0mg/l NAA+0.5mg/I BA supplemented MS medium. It is important to note that Cissus quadrangularis does not exhibit good callus growth in various concentrations of IAA or 2, 4-D etc. (16; 17). NAA uptake is enhanced and materialized to result in good callus growth at low concentration of bioactive Ca²⁺, which also helps in salt (Na⁺) exclusion and maintenance of primary metabolism. Cissus quadrangularis is susceptible to salt stress (as low as 50mM NaCl), yet callus growth can be maintained if it is raised on 3.0mg/l NAA+0.5mg/l BA in the presence of low concentration of Ca²⁺ (as in MS Medium). This growth alongwith improvement in the amounts of P and K+ are achieved as Ca2+ supposedly helps in binding and uptake of NAA (not of IAA or 2,4-D) alongwith activation of SOS pathway protecting the callus from injury. In this report we suggest the mechanism supporting bioactive Ca²⁺ and maintenance of uptake primary metabolism In vitro in Cissus quadrangularis.

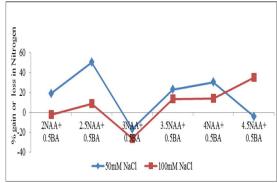


Fig.1: % gain or loss in Nitrogen (N) content in Cissus quadrangularis nodal calli on MS+PGRs+Salt supplemented media as compared to respective salt free control.

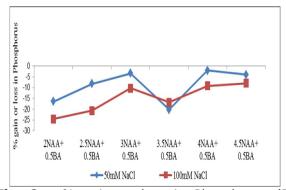


Fig. 2: % gain or loss in Phosphorus (P) content in *Cissus quadrangularis* nodal calli on MS+PGRs+Salt supplemented media as compared to respective salt free control.

Table.1: Effect of salt (NaCl) and Phytohormones (NAA & BA) on callus growth from nodal

explants in Cissus quadrangularis calli.

S.No.	MS + Growth	Salt	Callus	Morphology
	Regulators(mg/l)	(mM NaCl)	growth	of callus
1	2.0NAA+0.5BA	-	++	Loose, friable and brown
2	"	50mM	+	Loose, friable and brown
3	"	100mM	+	Loose, friable and brown
4	2.5NAA+0.5BA	-	++++	Loose, friable and brown
5	"	50mM	++	Loose, friable and brown
6	"	100mM	+	Loose, friable and brown
7	3.0NAA+0.5BA	-	+++	Loose, friable and brown
8	"	50mM	++	Loose, friable and brown
9	"	100mM	+	Loose, friable and brown
10	3.5NAA+0.5BA	-	++	Loose, friable and brown
11	"	50mM	+	Loose, friable and brown
12	"	100mM	+	Loose, friable and brown
13	4.0NAA+0.5BA	-	++	Loose, friable and brown
14	"	50mM	+	Loose, friable and brown
15	"	100mM	+	Loose, friable and brown
16	4.5NAA+0.5BA	-	++	Loose, friable and brown
17	"	50mM	+	Loose, friable and brown
18	II	100mM	+	Loose, friable and brown

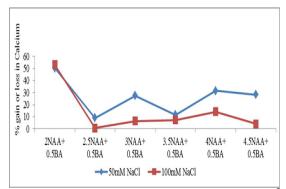


Fig.3: % gain or loss in Calcium (Ca²⁺) content in *Cissus quadrangularis* nodal calli on MS+PGRs+Salt supplemented media as compared to respective salt free control

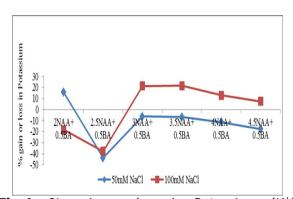


Fig.4: % gain or loss in Potassium (K⁺) content in *Cissus quadrangularis* nodal calli on MS+PGRs+Salt supplemented media as compared to respective salt free control

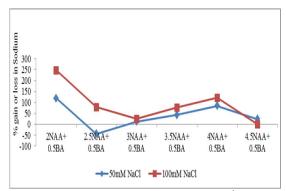


Fig.5: % gain or loss in Sodium (Na⁺) content in *Cissus quadrangularis* nodal calli on MS+PGRs+Salt supplemented media as compared to respective salt free control

Acknowledgement

One of the authors (Renu Rani) is grateful to Council of Scientific and Industrial Research for giving financial assistance in the form of Junior Research Fellowship.

References

- Garg B, Gupta IC, Saline Wastelands Environment and Plant Growth. Scientific Publishers (Jodhpur, India), 1997, pp 287.
- Mer RK, Prajith PK, Pndya DH, Pandey AN, Effect of salts on germination of seeds and growth of young plants of Hordeum vulgare, Triticum aestivum, Cicer arietinum and Brassica juncea. Journal of Agronomy and Crop Science 2000,185, 209-217.
- Bernstein L, Salt affected soils and plants. In 'Proceedings of the Paris Symposium, UNESCO (May1960)'. Arid Zone Research 1962, 18, 139-174.
- 4. Ramoliya PJ, Patel HM Pandey AN, Effect of salinization of soil on growth and macro –and micro–

- nutrient accumulation in seedlings of *Acacia catechu* (Mimosaceae). *Annals of Applied Biology* 2004, 144, 321-332.
- 5. Munns R, Physiological processes limiting plant growth in saline soils: some dogmas and hypotheses. *Plant Cell and Environment* 1993, 16, 15-24.
- Panthong A, Supraditaporn W, Kanjanapothi D, Taesotikul T, Reutrakul V, Analgesic, antiinflammatory and venotonic effects of Cissus quadrangularis. J. Ethnopharmacol 2007, 110, 264-270.
- Kar A, Borthakur SK, Wild vegetables of karbianglong district, Assam. Nat. Prod. Radiance 2008, 7, 448-460.
- 8. Meher A, Agrahari AK, Pradhan AR, Indian medicinal plant *Cissus quadrangularis* Linn. An ethnobotanical and ethnomedicinal review. *Herbal. Tech. Ind.* 2010, 15-17.
- 9. Murthy KNC, Vanitha A, Swamy MM, Ravishankar GA, Antioxidant and antimicrobial activity of *Cissus quadrangularis* L. *J. Med. Food* 2003, 6, 99-105.
- 10. Murashige T, Skoog FA, revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiol. Plantarum* 1962, 15, 473-497.
- 11. Snell DF, Snell CT, Colorometric methods of analysis including photometric methods. 1967, IV A 331-332.

- Olsen S, Cole C, Watanabe F, Dean L, Estimation of available phosphorus in soils by extraction with sodium bicarbonate. USDA Circular Nr 939, US Gov. Print. Office, Washington, D.C. 1954
- Pratt D, Fathi-Etti RA, Variation in organic and inorganic mineral components in young Eucalyptus seedlings under salinity stress. *Physical Plant* 1990, 79, 479-486.
- Zhu JK, Salt and drought stress signal transduction in plants. Annual Review of plant Biology 2002, 53, 247-273.
- Yamagami M, Haga K, Napier MN, Moritoshi lino, Two Distinct Signaling Pathways Participate in Auxin-Induced Swelling of Pea Epidermal Protoplasts. *Plant Physiol* 2004,134(2) 735-747.
- Sharma N, Nathawat RS, Gour K, Patni V, Establishment of callus tissue and effect rof growth regulators on enhanced sterol production in Cissus quadrangularis L. International Journal of Pharmacology 2011, 7, 653-658.
- 17. Teware K, Singh P, Mehta R, *In vitro* callus induction from stem explants of *Cissus quadrangularis* L. *International J. of Ayurvedic and Herbal Medicine* 2012, 2(1) 135-138.

Source of support: Nil
Conflict of interest: None Declared